

Lipid Peroxidation Inhibitory and Cytotoxic Activities of Two *Camellia* Species Growing Wild in Vietnam

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 and William N. Setzer^{7,8}

Abstract

Background: Leaves, flowers and young shoots of yellow *Camellia* species are often used as a substitute for green tea in Vietnam. Yellow *Camellia* species contain high levels of polyphenol compounds, and have shown activities such as antioxidant, anticancer, and cytotoxic.

Objectives: We qualitatively screened phytochemicals, quantified total polyphenols of 70% ethanol (EtOH) extracts of leaves, young shoots, and flowers of *Camellia vuquangensis* and *Camellia hatinhensis* in Ha Tinh Province, Vietnam. Furthermore, these extracts have been evaluated for lipid peroxidation inhibitory activities at the *in vivo* level in the BALB/c mouse model, and *in vitro* cytotoxic activities.

Materials and Methods: Chemical methods and thin-layer chromatographic (TLC) analysis were used for the qualitative screening of phytochemicals. High-performance liquid chromatography (HPLC) was used to determine total polyphenols. Cell survival was determined through optical density (OD) measured when the protein composition of the cells was stained with sulforhodamine B (SRB). Inhibition of lipid peroxidation was evaluated by determining the content of malondialdehyde (MDA), which is a product of membrane lipid peroxidation.

Results: The total polyphenol contents of all parts of the two species were comparable to that of green tea with values between 319.3 and 342.6 mg gallic acid equivalents (GAE/g) dry weight. Extracts of leaves, flowers of *C. vuquangensis* and *C. hatinhensis* showed strong lipid peroxidation inhibitory activities (IC₅₀: 7.92–17.45 µg/mL) and moderate cytotoxic activities against cell lines HepG2, A549, MCF7, SK-Mel-2, HT-29, and AGS (IC₅₀: 34.73–80.58 µg/mL).

Conclusion: These two tea species may be considered as herbal teas with various health benefits.

Keywords

Antiproliferative, *Camellia hatinhensis*, *Camellia vuquangensis*, Polyphenolics, Yellow *Camellia*.

Introduction

Lipid peroxidation is the cause of membrane damage including assembly, composition, structure, and dynamics of lipid membranes.¹ The degradation of products that are produced during peroxidation can indirectly damage other components of the cell.² Brain oxygen consumption during adenosine triphosphate (ATP) synthesis produces a high quantity of reactive oxygen species (ROS) as a byproduct, and biological cell membranes contain polyunsaturated fatty acids (PUFAs), which are easily peroxidized.³ Previous studies confirmed lipid peroxidation as the cause of diseases of the brain and the central nervous system.¹ In addition, peroxidation is involved in the pathogenesis of other human diseases such as ischemia-reperfusion injury, various types of cancers, Alzheimer's disease, rheumatoid arthritis, renal dysfunction, ocular degeneration, and diabetes mellitus.²

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Cancer is a global disease, the latest WHO report recorded 18.1 million new cases of cancer and 9.6 million deaths worldwide.⁴ Breast cancer is the leading cause of cancer death for women.⁵ In 2020, there were 685,000 deaths globally out of a total of 2.3 million women diagnosed with breast cancer.⁶ The incidence of breast cancer in men accounts for 0.5%–1% of all cases.⁶ Liver cancer is the most common cancer globally, accounting for 9.1% of all cancer deaths, with over 80% of diagnosed cases in underdeveloped countries.⁷ Globally, lung cancer is the most common cancer with 1.2 million new cases annually, responsible for 17.8% of all cancer deaths.⁸ Every year there are about 1.2 million new cases of colon cancer and about 600,000 deaths.⁹ In 2018 Gastric cancer was the cause of 784,000 deaths, and it is estimated that there are about 1 million new cases every year.¹⁰ About 2–3 million new cases are diagnosed with non-melanoma skin cancers, of which at least 132,000 malignant melanomas are diagnosed globally each year.¹¹

Camellia Linnaeus (1753) has between 120 and 280 species, distributed mainly in Asia; Vietnam and China are the two countries that are recording the largest number of species.¹² Most of the studies on chemical composition and biological activities have focused on three species with high economic value, namely *Camellia sinensis* (L.) Kuntze, *Camellia japonica* L., and *Camellia oleifera* Abel due to products such as tea and seed oil.¹² The secondary metabolites present in *C. sinensis* are mainly alkaloids, steroids, terpenoids, saponins, and polyphenols. Studies have shown that *Camellia* species exhibit activities such as antioxidant, antimicrobial, antifungal, antiviral, antitumor, and other health benefits.¹³ This study aims to qualitatively investigate phytochemical constituents, flavonoids, coumarins, tannins, saponins, polysaccharides, amino acids, alkaloids, quantification of epigallocatechin gallate (EGCG), and total polyphenols, lipid peroxidation inhibitory and cytotoxic activities of 70% ethanol extracts from different parts of two *Camellia* species growing wild in Vietnam. The results of this study may be used to compare with previously studied *Camellia* species, and to guide further studies on these two species.

Materials and Methods

Plants

Fresh shoots (leaf bud including two youngest leaves, NVQ), young leaves (third to fifth leaves, LVQ) and flowers (HVQ) of *Camellia vuquangensis* Luong, Tran and L. T. Nguyen were collected at Vu Quang National Park, Ha Tinh Province (18°13'27.290388"N; 105°20'20.08428"E; 130 m). Fresh shoots (leaf bud including two youngest leaves, NHT), young leaves (third to fifth leaves, LHT), and flowers (HHT) of *Camellia hatinhensis* Luong, Tran and L. T. Nguyen were collected at Vu Quang National Park, Ha Tinh Province

(18°18'57.92472"N; 105°20'47.51736"E). The fresh materials were washed with tap water and dried at 60°C for 2 days.

Animals and Cell Lines

Purebred BALB/c mice were provided by Biological Laboratory, Institute of Biotechnology—Vietnam Academy of Sciences. Healthy mice, disease-free, with experimental weights of 23–26 g at 12 weeks old were used for the Lipid Peroxidation inhibition test. The cancer cell lines Michigan Cancer Foundation-7 (MFC-7), human lung carcinoma (A549), human hepatocellular carcinoma (HepG2), human colorectal adenocarcinoma (HT-29), human melanoma (SK-Mel-2), and human gastric adenocarcinoma (AGS) were provided by Prof. Chi-Ying Huang, National Yang Ming Chiao Tung University, Taiwan.

Chemicals

Trolox, thiobarbituric acid (TBA), EGCG, galic acid, and ellipticine were purchased from Sigma-Aldrich. Acetonitrile and phosphoric acid were purchased from Merck. DMEM (Dulbecco's Modified Eagle Medium) supplemented with l-glutamine was purchased from Invitrogen (MA, USA). All other chemicals and reagents were of analytical grade.

Extracts Preparation

Dried materials were milled to a size of 3 mm, then soaked with methanol (MeOH) (Singapore) three times for 7 days each. Methanol extracts were evaporated with an Eyela N-1300V-W (Japan) at 50°C until they were completely dried, and stored at 4°C until further analysis.

Phytochemical Screening of Extracts

Accurately-weighed 100 mg samples of extracted residues were added to 25 mL of 70% ethanol, sonicated for 15 min, filtered, and the filtrates were used for *in vitro* qualitative reactions.

Test for Flavonoids

Shinoda's Test (Cyanidin Reaction)

Two milliliters of each filtrate was placed into a test tube, metallic magnesium powder was added, and then a few drops of concentrated hydrochloric acid were added. The test tube heated in a water bath for a few minutes and a purple-red color appears (positive reaction).¹⁴

Ferric Chloride Test

Two milliliters of each filtrate was placed into a test tube, then 3 drops of FeCl₃ 5% were added. A green-blue coloration indicated the presence of a phenolic hydroxyl group.¹⁴

Sodium Hydroxide Test

One milliliter of each filtrate was placed into a test tube, then 3 drops of NaOH 10% were added. A yellow coloration indicated the presence of flavonoids.¹⁴

Test for Flavonoids by Thin-Layer Chromatography (TLC)

Accurately-weighed 100 mg samples of extracted residues were added to 10 mL of distilled water in a glass beaker, sonicated for 10 min, then shook with 5 mL of *n*-butanol. The *n*-butanol extracts were evaporated with an Eyela N-1300V-W at 50°C until they were completely dried. The extracted residues were dissolved in 1 mL of methanol for chromatographic analysis. TLCs were developed for 20 min on 10 × 10 cm Silica gel 60 GF₂₅₄ (Merck, Germany) with ethyl acetate–acid formic–methanol–water (25:1:1:1, v:v:v:v) mobile phase. After growing the mobile phase to 8 cm, the TLCs were solvent-dried, sprayed with boric acid/oxalic acid reagent, and dried until the spots were visible. TLCs were observed under ultraviolet light at 366 nm, the chromatogram obtained with the test solution should have brighter luminescence spots than the chromatogram obtained with the test solution before the reagent was sprayed. The boric acid/oxalic acid reagent was prepared by mixing 10% boric acid solution and 10% oxalic acid solution in a ratio of 2:1 (v/v).¹⁵

Test for Coumarins

Lactone Ring Opening/Closing Reaction

One milliliter of the filtrate in 2 test tubes, into the first tube was added 0.5 mL of 10% NaOH solution, the second tube was not. After both test tubes were heated in a boiling water bath for several minutes, first tube with a yellow color appears. Then, adding 2 mL of distilled water to each tube, the first tube was clearer than the second, but after they were acidified with a few drops of concentrated HCl, both tubes were equally turbid (positive reaction).¹⁶

Reaction with Diazo Reagent

One milliliter of the filtrate was added to the test tube, then 2 mL of 10% NaOH solution was added to the test tube. Test tubes were heated to boiling, cooled, added a few drops of diazo reagent (freshly prepared), brick red precipitate was observed (positive reaction). Diazo reagent was prepared according to the following protocol. 0.90 g of sulfanilic acid was dissolved in 9 mL of concentrated hydrochloride acid by heating, then diluted with water to 100 mL. Next, 10 mL of the above solution was soaked in ice, then 10 mL of pre-cooled 4.5% sodium nitrite solution (by immersion in ice) was added. The mixture was soaked in ice water for about 15 min, after which 20 mL of 10% sodium carbonate solution was added.

Test for Coumarins by TLC

Ten milliliters methanol was used to dissolve about 10 mg extracted residue, sonicated for 5 min, filtered, obtained TLC analytical solution. TLCs were developed for 20 min on 10 × 10 cm Silica gel 60 GF₂₅₄ (Merck, Germany) with toluene–ethyl acetate–acetone–formic acid (5:2:2:1, v:v:v:v) mobile phase. After growing the mobile phase to 8 cm, the TLCs were solvent-dried, sprayed with 2% KOH/ethanol reagent, and dried until the spots were visible. TLCs were observed under ultraviolet light at 366 nm, the chromatogram obtained with the test solution should have brighter luminescence spots than the chromatogram obtained with the test solution before the reagent was sprayed.¹⁵

Test for Tannins

The First Test Tube

Two milliliters of filtrate was put into the test tube, then 2 drops of 5% FeCl₃ solution were added. The reaction is positive when a greenish-black or light-brown precipitate is present.¹⁴

The Second Test Tube

Two milliliters of filtrate was put into the test tube, then 2 drops of 10% Pb(C₂H₃O₂)₂ solution were added. The reaction is positive when a precipitate is present.

The Third Test Tube

Two milliliters of filtrate was put into the test tube, then 5 drops of 1% gelatin solution were added. The reaction is positive when a white precipitate appears.

Test for Saponins

Observation of Foaming

One milliliter of the filtrate was put into a test tube of 20-mL capacity, to which 10 mL of distilled water was added. Test tubes were thumb-cap, shaken vigorously vertically for 5 min, then left to stand still and observe. The reaction is positive when the foam is stable after 10 min.¹⁴

Salkowski's Test

One milliliter of the filtrate was added to the test tube, and the filtrate was evaporated to obtain residue. The residue was dissolved in 1 mL of acetic anhydride, then 0.5 mL of chloroform was added to the solution. One to two milliliters of H₂SO₄ was slowly dripped into the wall of the test tube with a pipette. The reaction is positive when a purple-red ring appears on the interface.¹⁷

Test for Saponins by TLC

Ten milliliters methanol was used to dissolve about 10 mg extracted residue, sonicated for 5 min, filtered, obtained TLC

analytical solution. TLCs were developed for 20 min on 10 × 10 cm Silica gel 60 GF₂₅₄ (Merck, Germany) with chloroform–methanol–water (65:35:10, v:v:v) mobile phase. After growing the mobile phase to 8 cm, the TLCs were solvent-dried, sprayed with 10% H₂SO₄/ethanol reagent, and dried until the spots were visible. TLCs have been observed under normal light, the chromatogram of the test solution should have pink, purple spots.¹⁵

Standard Test for free Reducing Sugar (Fehling's Test)

One milliliter of the filtrate was added to the test tube. Then 0.5 mL Fehling A and 0.5 mL Fehling B were added to the test tube. Test tubes were boiled in a water bath for several minutes until a brick red precipitate appeared (positive reaction). Fehling A reagent was prepared by dissolving 7.0 g CuSO₄·5H₂O in distilled water containing 2 drops of dilute sulfuric acid solution. Fehling B reagent was prepared by dissolving 35 g of potassium tartrate (C₄H₄K₂O₆) and 12 g of NaOH in 100 mL of distilled water.^{14,17}

Test for Polysaccharides (Iodine Test)

The first test tube was composed of 4 mL of distilled water and 5 drops of Lugol reagent. The second test tube was composed of 4 mL of filtrate and 5 drops of Lugol reagent. Observed that the color of second test tube is darker than first test tube, the reaction is positive. Lugol reagent was prepared according to the following protocol. Ten gram of potassium iodide (KI) dissolved in about 20–30 mL of distilled water. Next, 5 g of iodine was added and gently heated, with constant stirring, until the iodine was dissolved, then diluted with distilled water to 100 mL to obtain reagent Lugol. Store in amber sealed glass bottle in a dark place.¹⁸

Test for Amino Acids

One milliliter of the filtrate was put into the test tube, then a few drops of 3% ninhydrin reagent was added, boiled the test tube for about 2 min in a water bath, the solution in the test tube turned purple (reactive, positive response).¹⁹

Test for Alkaloids

One hundred milligrams of extracted residue was put into a 50 mL conical flask, 15 mL of 2% H₂SO₄ solution was added, then boiled for a few minutes. The solution was cooled, the extract was filtered into a decanter, the filtrate alkalized with 6N NH₄OH solution to alkaline pH. The alkaloids were extracted with chloroform (CHCl₃) three times, 5 mL each. The CHCl₃ extract was pooled and shaken with 2% H₂SO₄ solution. Acidic water layer was added to 3 test tubes, each tube was about 1 mL and the following reactions were carried out:

1. Mayer reagent was added to the first test tube.
2. Mayer reagent was prepared by dissolving a mixture of 1.36 g of mercuric chloride (HgCl₂) and 5 g of potassium iodide (KI) in 100 mL of water.
3. Bouchardat reagent was added to the second test tube.
4. Bouchardat reagent was prepared by dissolving 2 g of iodine and 4 g of potassium iodide in 10 mL of distilled water, after the iodine had dissolved, add enough water to make 100 mL.
5. Dragendorff reagent was added to the third test tube.

The Dragendorff reagent was prepared according to the following protocol: 0.5 g of Bismuth nitrate was dissolved in 10 mL of concentrated hydrochloric acid, then mixed in a 1:1 (v/v) ratio with a solution containing 4 g of KI in 10 mL of distillate.¹⁷

Test for Alkaloids by TLC

One hundred milligrams of extracted residue was put into a 50 mL conical flask, 15 mL of 2% H₂SO₄ solution was added, then boiled for a few minutes. The solution was cooled, filtered into a decanter, alkalized with 6N NH₄OH solution to alkaline pH. The alkaloids were extracted with chloroform (CHCl₃) three times, 5 mL each. The CHCl₃ extracts were combined, completely evaporated, residue was dissolved in 2 mL of methanol, and the obtained solution was used for analysis by TLC. TLCs were developed for 20 min on 10 × 10 cm Silica gel 60 GF₂₅₄ (Merck, Germany) with chloroform–methanol–NH₄OH solution with concentration of 6N (7:1:0.1) mobile phase. After growing the mobile phase to 8 cm, the TLCs were solvent-dried, sprayed with Dragendorff reagent. The TLCs were observed under normal light, the chromatogram of the test solution should show yellow-orange spots.¹⁵

Qualitative Analysis of Epigallocatechin Gallate (EGCG) from Extracted Residues Using High-Performance Liquid Chromatography (HPLC)

Five milligrams extracted residue was accurately weighed and completely dissolved with 10 mL methanol under ultrasonic wave conditions. One milligram of EGCG was accurately weighed, then completely dissolved with 10 mL of methanol under ultrasonic conditions. Samples were filtered through a 0.45 μm filter before chromatography.

Samples were analyzed for EGCG by HPLC containing Agilent C₁₈ column (5 μm particle size, 4.5 – 250 mm). Ten microliters of each sample were injected and the column temperature was 40°C. The 0.1% acetonitrile–phosphoric acid mobile phase (ratio 10/90) was used with a flow rate of 1 mL/min. Qualitative analysis of EGCG was carried out by ultraviolet (UV) detector at 270 nm.¹⁵

Quantitative Analysis of Total Polyphenols

Prepare of Standard Solution

Fifty milligrams of standard gallic acid was accurately weighed into a 100 mL brown volumetric flask, and enough water was added to the mark for dissolution. Five milliliters of the above solution was accurately aspirated into a brown 50 mL volumetric flask, just enough water was added, and the solution was shaken to obtain a solution with a concentration of gallic acid about 0.05 mg/mL.

Preparation of Test Solution

One hundred milligrams extracted residue was accurately weighed into a 25 mL brown volumetric flask, 20 mL of water was added and ultrasound for 15 min, added just enough water, and mixed well. The solution was filtered and the first 10 mL of the filtrate was discarded.

Investigation of optical absorption: 1.6 mL of standard solution and 0.8 mL of test solution were accurately aspirated into a brown 10 mL volumetric flask; then 0.4 mL of Folin-Ciocalteu reagent, 4.0 mL of distilled water and Na_2CO_3 solution were added to each flask to make up to 10 mL, and shaken well. The UV absorption spectrum of the analytical solutions were scanned in the wavelength range from 400 to 850 nm, the maximum absorption wavelength was selected for analysis.

Constructing a Calibration Curve for Gallic Acid

The gallic acid standard solution was diluted in series of solutions S1, S2, S3, S4, S5, and blank solution into separate brown 25 mL flasks with the following formulas (Table 1):

Table 1. Preparation of Gallic Acid Calibration Solutions.

Composition	S1	S2	S3	S4	S5	Blank
Standard solution (mL)	1.0	2.0	3.0	4.0	5.0	0
Folin-Ciocalteu reagent (mL)	1	1	1	1	1	1
Distilled water (mL)	11	10	9	8	7	12
Solution of 29% Na_2CO_3 (mL)	Total volume is just enough 25 mL					

The absorbances of the solutions were determined and a calibration curve was constructed with absorbance as the vertical axis and the solution concentration as the horizontal axis.

Quantitative Analysis of Total Polyphenols

The 0.8 mL of test solution was accurately aspirated into a brown 10 mL volumetric flask, then 0.4 mL of Folin-Ciocalteu reagent, 4.0 mL of distilled water and Na_2CO_3 solution were added to make up 10 mL, and was shaken well. The absorbances of the solutions were measured. The concentration of the photometric analytical solution of the test sample is calculated by the formula:

$$C = \frac{Y-b}{a} \times \frac{H}{100} (\mu\text{g/mL})$$

where C is the concentration of the photometric solution of the test sample ($\mu\text{g/mL}$); A is the absorbance of the test solution; a, b are the coefficients of the standard curve equation for gallic acid; H is the purity of the standard substance (%).

The polyphenol content in the extracted residue sample was determined according to the formula:

$$X_1 = \frac{C \times 25 \times 25}{m} (\text{mgGAE/g})$$

where X is the total polyphenol content in extracted residue samples calculated by gallic acid (mg GAE/g); C is the concentration of the photometric analytical solution of the test sample ($\mu\text{g/mL}$); m is the mass of extracted residue (mg).

Cytotoxic Activity Assay

Cell Culture

The cancer cell lines were cultured according to the routine guidelines of ATCC (American Type Culture Collection).²⁰ Specifically, six experimental cancer cell lines were grown in culture medium DMEM supplemented with 2 mM L-glutamine and fetal bovine serum (10% FBS) and other necessary components at 5% CO_2 condition, at 37°C. All cell lines (MCF-7, A549, HepG2, HT-29, SK-Mel-2, and AGS) were subcultured after three days.

Process subculture was performed as follows: first, the possibility of development of the cell line was examined by the microscope. The cells were washed with PBS to remove the original culture medium. Trypsinization with trypsin-ethylenediaminetetraacetic acid (EDTA) was carried out at 37°C for 1–3 min to separate the adherent cells, then 5 mL of culture medium was added to stop the effect of trypsin. The suitable subculture ratio was determined by cell density counting using an erythrocyte counting chamber. Trypsin was removed by centrifugation at 9000–1000 rpm for 5 min, then the solution was removed and the residue was retained. The cell residue was diluted with the necessary culture medium to obtain a cell density suitable for cell growth. The culture flasks containing the cells were maintained in the culture chamber at 37°C and 5% CO_2 . During culture, for non-adherent cell types, trypsinization was not performed.

Cytotoxic Activity Assay for Monolayer Cultured Cells

The cytotoxic activities were performed according to the method of Monks (1991).²¹ The test was carried out to

determine the total cellular protein content based on the optical density (OD) measured when the protein composition of the cells was stained with sulforhodamine B (SRB). The more cells (more protein) the greater the OD value because the measured OD value is directly proportional to the amount of SRB bound to the protein molecule.

The solution of extracted residue or pure compound (10 μ L) was diluted in 10% dimethyl sulfoxide (DMSO), then put into a 96-well tray to achieve a series of concentrations of 100 μ g/mL, 20 μ g/mL, 4 μ g/mL, 0.8 μ g/mL, and 0.16 μ g/mL. Cells were separated by trypsinization, then counted in a counting chamber to adjust the density to suit the assays. The test wells were added to the appropriate amount of cells (in 190 μ L of medium). Another 96-well tray containing no extracted residue or pure compound but containing cancer cells (180 μ L) was used as a day 0 control. After 1 h, in the day 0 control tray the cells were fixed with trichloroacetic acid (TCA). After the growth period in the CO₂ incubator, cells were fixed to the bottom of the wells with TCA for 1 h and were stained with 0.4% SRB for 30 min at 37°C. SRB was removed from the test wells, then the test wells were washed three times with acetic acid and then air-dried at room temperature. The bound SRB was dissolved using 10 mM unbuffered Tris base, then the protein molecules were stained, put on a plate shaker and gently shaken for 10 min and used an ELISA Plate Reader (Bio-Rad) to read the results of the color content of the SRB dye through the absorption spectrum at 515 nm. The viability of cells in the presence of extracted residue or pure compound was determined by the following formula:

$$\text{Percentage of cell viability (\%)} = \frac{[\text{OD (reagent)} - \text{OD (day 0)}] \times 100}{\text{OD (negative control)} - \text{OD (day 0)}}$$

The assays were repeated three times to ensure accuracy. Ellipticine was used as a positive control and tested at concentrations of 10 μ g/mL, 2 μ g/mL, 0.4 μ g/mL, and 0.08 μ g/mL. The 10% DMSO solution was used as the negative control.

Lipid Peroxidation Inhibitory Activity (MDA Assay)

The antioxidant activity assay through inhibition of membrane lipid peroxidation (MDA assay) was performed according to the protocol previously described with slight variation.^{21–23} The ability of the sample to inhibit lipid peroxidation was determined by determining the content of malondialdehyde (MDA), which is a product of membrane lipid peroxidation. MDA is able to react with TBA to form trimethin complex (pink color) with maximum absorption peak at $\lambda = 532$ nm.

All experimental procedures that involved mice were conducted in accordance with the “Guideline for the Care and Use of Laboratory Animals” which was approved by the Institute of Biotechnology—Vietnam Academy of Science and Technology.

Preparation of Test Sample

The extracted residue or pure compound was weighed accurately, then diluted to concentrations of 10,000 μ g/mL, 2000 μ g/mL, 400 μ g/mL, 80 μ g/mL, and 16 μ g/mL. Then 0.1 mL of test sample at the test concentrations was reacted with 1 mL of brain homogenate and added 0.8 mL of phosphate buffer, 0.1 mL of Fenton system, the final concentrations of the sample in the test tube were reduced 20 times to 500 μ g/mL, 100 μ g/mL, 20 μ g/mL, 4 μ g/mL, and 0.8 μ g/mL. The standard set of Fenton reagents we used for the experiments including ammonium iron (II) sulfate hexahydrate [(NH₄)₂Fe(SO₄)₂·6H₂O] (Sigma Ultra, minimum 99% pure); 0.5 M EDTA solution, pH 8.0 (Ambion); (+) Sodium L-ascorbate (Sigma), 30% hydrogen peroxide (Sigma Aldrich), thiourea (Sigma), and H₂O.

MDA Assay

Mouse brain was separated and crushed homogeneous in phosphate buffer solution (pH = 7.4) in the ratio 1:10 at temperature 0–4°C. One milliliter (1 mL) of brain homogenate was added and 0.1 mL of test sample at different concentrations, 0.8 mL of phosphate buffer and 0.1 mL of Fenton reagent (FeSO₄ 0.1 mM:H₂O₂ 15 mM in the ratio 1:1). The mixture was incubated at 37°C for 15 min. One milliliter (1 mL) of 10% trichloroacetic acid was added to stop the reaction, then centrifuged at 12,000 rpm for 5 min. The clear solution was reacted with 1 mL of 0.8% TBA (in a ratio of 2:1) and incubated at 100°C for 15 min. After cooling, the absorbance was measured at $\lambda = 532$ nm. Trolox was used as a positive control. Percentage inhibition was calculated with the formula:

$$\text{HTCO(\%)} = \frac{(\text{OD}_C - \text{OD}_T) \times 100}{\text{OD}_C}$$

where OD_C is the optical density of the control well without the test sample; OD_T is the optical density of the test specimen.

Statistical Analysis

The IC₅₀ values (concentration that inhibits 50% of cell growth) were determined using Table-Curve 2D computer software, version 4.0. ANOVA testing was performed using Minitab version 19 software.

Table 2. Yield Extract of Plant Material Samples.

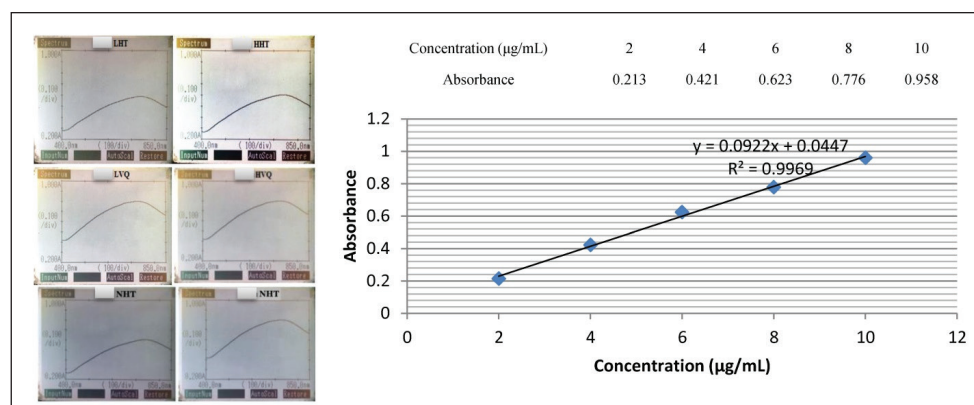
No.	Code	Volume of Liquid Extract (g)	Volume of Extracted Residue (g)	Moisture of Extracted Residue (%)
1	LHT	10.75	3.68	16.55
2	LVQ	3.80	2.10	5.05
3	HHT	3.7	2.79	12.70
4	HVQ	3.5	2.45	12.40
5	NHT	3.7	2.50	11.60
6	NVQ	4.5	3.25	11.30

Table 3. The Results of Phytochemical Screening of Extracted Residues.

Code	Flavonoids	Coumarins	Tannins	Saponins	Reducing Sugars	Polysaccharids	Amino Acids	Alkaloids	EGCG
LHT	++	–	+	+	++	++	–	–	–
LVQ	++	–	–	+	++	++	–	–	–
HHT	++	–	+	+	++	++	–	–	–
HVQ	++	–	–	++	++	++	–	–	–
NHT	++	–	–	++	++	++	–	–	–
NVQ	++	–	–	++	++	++	–	–	–

Note: (–): negative, (+): positive, (++): clearly positive.

Abbreviation: EGCG, epigallocatechin gallate.

**Figure 1.** Absorption Peak Values of Extracted Residues and Calibration Curve of Standard Gallic Acid.

Results

Yield of the Extraction Processes

The yields of the studied samples are presented in Table 2. The yields of the studied samples were not significantly different and varied between 2.10 and 3.68 g.

Phytochemical Screening of Extracted Residues

The results of phytochemical screening of extracted residues are summarized in Table 3. None of the samples showed the presence of amino acids, alkaloids, coumarins or EGCG. Two

samples of LHT and HHT tested positive for tannins, while the other samples were negative.

Quantification of Total Polyphenols

The results of scanning the spectrum of standard solutions and test solutions on a spectrophotometer in the wavelength range from 300 to 800 nm have shown absorption peak values at wavelengths 737.0–740.0 nm. Therefore, 740.0 nm was selected as the analytical wavelength (Figure 1).

The result of the standard curve construction according to the gallic acid are presented in Figure 1.

Table 4. Total Polyphenol Contents in Extracted Residues (mg GAE/g).

Code	Volume of Extracted Residues (mg)	Absorbance	Total Polyphenol Content in Extracted Residues (mg GAE/g)	Mean $\pm$ SD(mg GAE/g)
LHT	10.7	0.554	322.66	322.61 ^a $\pm$ 1.70
	10.4	0.537	320.88	
	10.5	0.547	324.28	
LVQ	10.3	0.568	344.40	342.60 ^b $\pm$ 2.50
	10.5	0.577	343.65	
	10.8	0.586	339.75	
HHT	10.8	0.549	316.53	319.30 ^a $\pm$ 2.60
	10.5	0.543	321.70	
	10.1	0.521	319.67	
HVQ	10.4	0.578	324.42	329.57 ^a $\pm$ 4.67
	10.5	0.578	333.55	
	10.9	0.589	330.73	
NHT	10.2	0.529	312.53	322.21 ^a $\pm$ 8.69
	10.4	0.513	324.77	
	10.3	0.529	329.34	
NVQ	10.2	0.562	324.44	325.94 ^a $\pm$ 3.32
	10.4	0.573	323.63	
	10.9	0.585	329.75	

Note: Results are means $\pm$ SD ($n = 3$). Values followed by the same letter (a,b) are not statistically different at $p < 0.05$ as measured by the Tukey's test.

Abbreviation: GAE, gallic acid equivalent; SD, standard deviation.

The quantitative results of total polyphenols in the extracted residues were between 319.30 and 342.60 mg GAE/g (Table 4). The total polyphenol contents of the samples HVQ, NVQ, LHT, NHT, and HHT were similar and all of them were lower than that of LVQ ($p = .001$).

Phenolic compounds are present in all the different parts of *Camellia* species (such as fresh green leaves, fresh tea buds, or flushes), and the total polyphenols content corresponding to the quality of the teas (Table 5).

Polyphenols are the main constituents in tea, accounting for 20%–30% of the dry weight of tea, of which catechins are the dominant group with 60%–80% of total polyphenols.⁴⁸ Table 5 shows that EtOH is the best solvent to extract polyphenols from *Camellia* species. The total polyphenols in the leaves, flowers, and buds of *C. vuquangensis* and *C. hatinhensis* ranged from 319.3 to 342.6 (mg GAE/g), which are comparable to the total polyphenols of different parts of *C. sinensis*,^{26,35,36} and lower than flowers of *Camellia nitidissima*.⁴⁴

EGCG is the most important polyphenol of *C. sinensis*,^{48,49} fresh bud and the first two leaves contain the highest amount which represents approximately 8–12% of dry weight.^{48–51} All parts (leaves, flowers, shoots) of *C. vuquangensis* and *C. hatinhensis* in this study did not have the presence of EGCG, galocatechin gallate (GCG), epicatechin gallate (ECG), or catechin gallate (CG) (Figure 2).

Cytotoxic Activity

The results of cytotoxicity assessment against cancer cell lines are summarized in Table 6. The samples HHT, LHT, HVQ showed moderate cytotoxic activity against different cancer cell lines HepG2, A549, MCF7, SK-Mel-2, HT-29, and AGS with IC_{50} values in the range 38.01–59.07 μ g/mL, 48.01–77.88 μ g/mL, 37.13–72.27 μ g/mL, 34.73–60.78 μ g/mL, 39.31–80.31 μ g/mL, and 36.89–80.58 μ g/mL, respectively. Meanwhile, sample LVQ showed weak activity with IC_{50} values > 100 μ g/mL for all cell lines that were tested. According to the standards of the US National Cancer Institute (NCI), the extract is considered to have good activity with $IC_{50} \leq 20$ μ g/mL, while the purified substance is considered to have good activity when $IC_{50} \leq 5$ μ M.⁵²

The cytotoxic activities of several species of *Camellia* that have been reported by other authors are summarized in Table 7, which showed stronger cytotoxic activity of *C. vuquangensis* and *C. hatinhensis* compared with other previously reported species. The cause of the strong cytotoxicity of *C. vuquangensis* and *C. hatinhensis* may be related to their high total polyphenols, but EGCG, GCG, ECG, and CG were not responsible.

The results of the determination of antioxidant activity through inhibition of membrane lipid peroxidation (MDA assay) are presented in Table 8. All samples LHT, LVQ, HHT,

Table 5. Total Polyphenols of Some *Camellia* Species that were Previously Reported.

Species	Parts	Solvent	Total Polyphenol/Phenolic(mg GAE/g)	Ref.
<i>Camellia sinensis</i>	Tender leaves	Nd	234.35	24
	Green tea ^a	70% MeOH	210.2–143.2	25
	Green tea ^a	70% MeOH	108.8–323.6	26
	Leaves	95% EtOH	355.26–457.89	27
	Leaves	100% MeOH	152–223	28
	Young shoots	70% MeOH	167.2–204.2	29
	Fresh two leaves and a bud	70% MeOH	266.33–266.49	30
	Leaves	H ₂ O	20.7–36	31
	Green tea ^b	H ₂ O	51.20	32
	Leaves and buds	50% MeOH	25.8–98.2	33
	Fresh leaves	70% MeOH	235.47	34
	Indoor-withered leaves	70% MeOH	203.47	34
	Solar-withered leaves	70% MeOH	158.45	34
	Green tea	H ₂ O	112.3	35
	Young expanding leaves	75% EtOH	329.395	36
	Buds	75% EtOH	346.219	36
	Leaves	60% EtOH	352.2	37
	Leaves	40% EtOH	104.03–290.09	38
	Shoots with a bud with two leaves	50% MeOH	173.90–201.04	39
	Leaves	70% MeOH	125.51–136.34	40
<i>Camellia oleifera</i>	Fruit hull	70% Acetone	42.52–234.90	41
<i>Camellia nitidissima</i>	Flowers	Dichloromethane fraction	85.02	42
		Ethyl acetate fraction	345.14	42
		<i>n</i> -Butanol fraction	164.19	42
		Water fraction	31.69	42
		95% EtOH	170.74	42
	Leaves	10% EtOH	34.47	43
	Flower	90% EtOH to 95% EtOH	550	44
	Leaves	H ₂ O	8.36	45
	Leaves	Acetone/ethanol/water (AEW; 2:2:1, v/ v/v)	152.2	46
	Leaves	H ₂ O	8.61	45
<i>Camellia nitidissima</i> var. <i>microcarpa</i>	Leaves	H ₂ O	10.30	45
<i>Camellia murauchii</i>	Leaves	H ₂ O	9.70	45
<i>Camellia impressinervis</i>	Leaves	Acetone/ethanol/water (AEW; 2:2:1, v/ v/v)	475.6	46
	Leaves	H ₂ O	9.70	45
<i>Camellia euphlebia</i>	Leaves	Acetone/ethanol/water (AEW; 2:2:1, v/ v/v)	257.8	46
	Leaves	H ₂ O	5.87	45
<i>Camellia tunghinensis</i>	Leaves	Acetone/ethanol/water (AEW; 2:2:1, v/ v/v)	67.2	46
	Leaves	H ₂ O	5.87	45
<i>Camellia microcarpa</i>	Leaves	Acetone/ethanol/water (AEW; 2:2:1, v/ v/v)	201.1	46
<i>Camellia chrysantha</i>	Leaves	Acetone/ethanol/water (AEW; 2:2:1, v/ v/v)	138.2	46
<i>Camellia ptilophylla</i>	Young shoots	MeOH	280.74	47
<i>Camellia kucha</i>	Young shoots	MeOH	234.08	47

Note: ^aCommercial tea leaf products in China. ^bApical bud and two or three adjoining leaves.

Abbreviations: GAE, gallic acid equivalent; Nd, not determined.

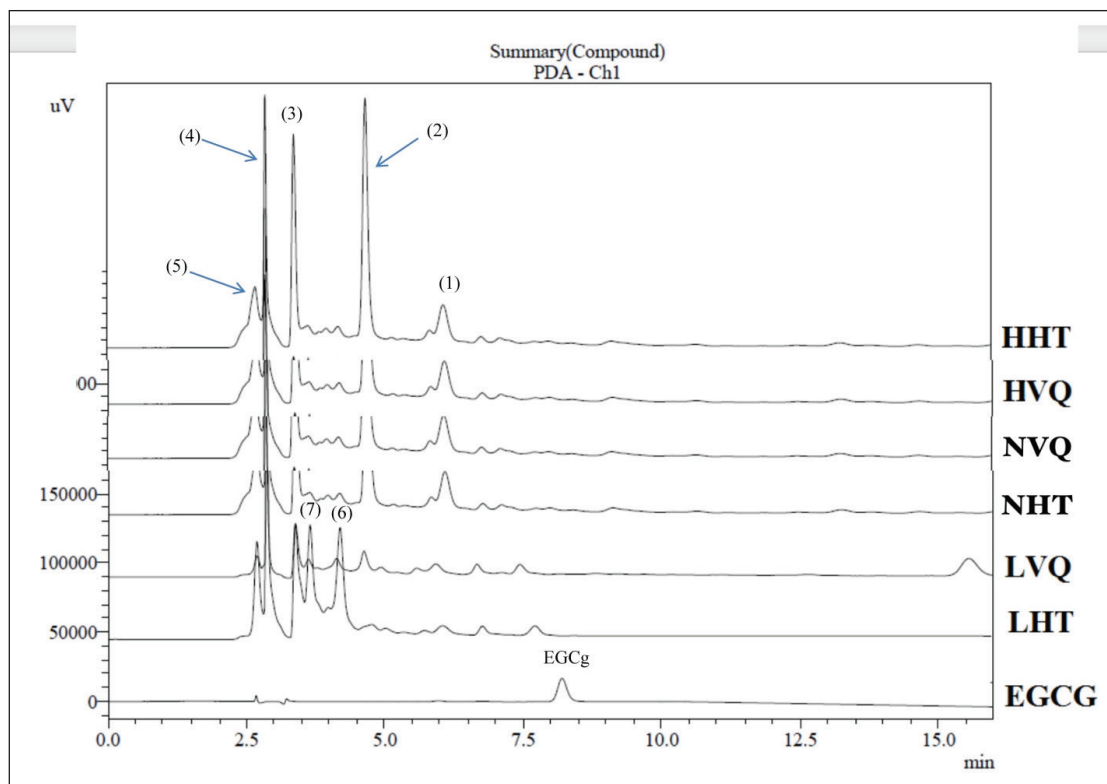


Figure 2. HPLC Chromatogram of Extracted Residues at 270 nm..

Table 6. Cytotoxicity of Extracted Residues (IC_{50} , $\mu\text{g/mL}$).

Code	HepG2	A549	MCF7	HT-29	SK-Mel-2	AGS
LHT	$59.07^a \pm 1.17$	$76.26^a \pm 1.68$	$72.27^a \pm 1.71$	$58.12^a \pm 1.74$	$56.22^a \pm 1.69$	$68.34^a \pm 2.27$
LVQ	$>100^b$	$>100^b$	$>100^b$	$>100^b$	$>100^b$	$>100^b$
HHT	$38.01^c \pm 2.02$	$48.01^c \pm 2.14$	$37.13^c \pm 1.66$	$39.31^c \pm 1.95$	$34.73^c \pm 1.86$	$36.89^c \pm 1.51$
HVQ	$56.11^a \pm 2.52$	$77.88^a \pm 1.93$	$61.57^d \pm 2.60$	$80.31^d \pm 1.82$	$60.78^d \pm 2.49$	$80.58^d \pm 1.48$
NHT	Nt	Nt	Nt	Nt	Nt	Nt
NVQ	Nt	Nt	Nt	Nt	Nt	Nt
Ellipticine	$0.39^d \pm 0.21$	$0.41^d \pm 0.21$	$0.38^e \pm 0.16$	$0.38^e \pm 0.13$	$0.42^e \pm 0.16$	$0.41^e \pm 0.13$

Note: Results are means $\pm$ SD ($n = 3$). For each column, values followed by the same letter (a–e) are not statistically different at $p < 0.05$ as measured by the Tukey's test.

Abbreviation: Nt, not tested.

and HVQ showed strong lipid peroxidation inhibitory activity comparable to positive control with IC_{50} values in the range 7.92–17.45 $\mu\text{g/mL}$.

Discussion

Jiang et al. analyzed the polyphenols in petals, stamens and sepals of the full flowering flowers of 22 species of yellow *Camellia* by HPLC and showed that EGCG was only a minor

component or was absent in many species.⁶⁵ For *Camellia piliphylla*, EGCG acted as a minor component in low concentrations.^{47,66} Our results are in agreement with published results by Wang et al. who reported on catechins and the bioactivity of *Camellia sasanqua*.⁶⁷ The leaves of the yellow *Camellia* species in the study of Lin et al. contained significantly high levels of EGCG.⁴⁵ EGCG has also been found with high content in the leaves of *Camellia assamica*,⁴⁶ *Camellia kucha*,⁴⁷ and *Camellia taliensis*.⁶⁸ The 60% ethanol

Table 7. Cytotoxicity of Some *Camellia* Species that were Previously Reported.

Plant	Part	Total Polyphenol/Phenolic	Cell	IC ₅₀ µg/mL
<i>C. sinensis</i>	Green tea	Nd	HepG2	72.0 ⁵³
	Green Tea	Nd	A549	112 ⁵⁴
	Catechin-rich green tea extract	Nd	MCF-7	324.5 ⁵⁵
	White tea	863 ± 5.6 mg GAE/g	HT-29	86.68 ⁵⁶
	Black Tea	341.8 mg GAE/g	AGS	264.3 ⁵⁷
<i>C. sinensis</i> var. <i>sinensis</i>	Black tea	678–710 mg GAE/L	A549	64.4–135.6 ⁵⁸
<i>C. euphlebia</i>	Flowers	5.11%	HepG2	>150 ⁵⁹
	Flowers	8.88%	HepG2	>150 ⁵⁹
<i>C. ptilophylla</i>	Leaves	Nd	HepG2	292 ⁶⁰
<i>C. sinensis</i> var. <i>assamica</i>	Leaves	1195–1786 mg GAE/100 g	HepG2	1280 ⁶¹
	Leaves	1195–1786 mg GAE/100 g	A549	399.8 ⁶¹
	Green tea	19.1 mg GAE/100 g	A549	450 ⁶¹
	Green tea	19.1 mg GAE/100 g	HT-29	780 ⁶¹
	Green tea	2700.56	HepG2	>1000 ⁶²
<i>C. japonica</i>	Flowers	Nd	MCF-7	>100 ⁶³
<i>C. tenuifolia</i>	Flowers	Nd	MCF-7	>100 ⁶³
<i>C. oleifera</i>	Flowers	Nd	MCF-7	>100 ⁶³
<i>C. sinensis</i>	Flowers	Nd	MCF-7	>100 ⁶³
<i>C. chrysantha</i>	Leaves	Mix polyphenols 40%	Hep G2	103 ⁶⁴
	Leaves	Mix polyphenols 40%	MCF7	107 ⁶⁴
	Leaves	Mix polyphenols 40%	SK-Mel-2	48 ⁶⁴

Abbreviations: GAE, gallic acid equivalent; Nd, Not determined.

Table 8. Lipid Peroxidation Inhibitory Activity of Extracted Residues.

Concentration (µg/mL)	Percentage of Lipid Peroxidation Inhibitory (%)				
	LHT	LVQ	HHT	HVQ	Trolox
100	77.49	77.49	82.09	77.57	76.15
20	58.46	68.29	73.74	60.30	71.85
4	23.43	34.96	31.71	32.63	48.00
0.8	18.40	24.84	19.39	24.70	16.31
IC ₅₀	17.45 ^a ± 1.26	8.15 ^b ± 1.01	7.92 ^b ± 0.57	12.29 ^c ± 1.12	7.04 ^b ± 0.56

Note: Results are means ± SD (n = 3). Values followed by the same letter (a–c) are not statistically different at p < 0.05 as measured by the Tukey's test.

Abbreviation: SD, standard deviation.

extracts of the leaves of *C. japonica* showed α- and β-anomers of pedunculagin, catechin, and epicatechin (EC).⁶⁹

The LVQ sample exhibited the weakest cytotoxic activities, which may have been related to the absence of components corresponding to the peaks (1) and (2), at the same time with a low content of the component corresponding to the peaks (3) and (5) (Figure 2). The high content of components corresponding to peaks (2) and (3) may have been responsible

for the strong cytotoxic activities of the HHT sample. The cytotoxic activities of the LHT sample may have been related to the components with peaks (6) and (7), respectively. Many studies have shown that catechin compounds were not primarily responsible for the cytotoxic activities, rather, flavonoid glycosides may have played a major role.^{45,58,59,62,63}

MDA is an indicator of lipid peroxidation. The concentrations of MDA in the liver, brain, and blood of the

rats treated with green tea (EGCG (337 mg/L), epigallocatechin (EGC) (268 mg/L), EC (90 mg/L), ECG (60 mg/L), and caffeic acid (35 mg/L) were significantly reduced compared with the control group, especially in the brain, which was decreased more than two times.⁷⁰ Green tea catechin polyphenols reduced MDA levels in the cerebral cortex of rats with intermittent hypoxia by 40%.⁷¹ Total flavones of *Camellia* at doses of 40 and 80 mg/kg reduced MDA level in brain tissues.⁷² The MDA was significantly reduced in the brains of mice treated with gallic acid and green tea leaf extract.⁷³ The 95% ethanol extract obtained from fresh leaves of *C. sinensis* at a dosage of 200 mg/kg significantly reduced serum and hepatic MDA concentration in rats.⁷⁴ Ethanolic extracts obtained from fresh flowers of *C. japonica* at doses of 40, 80 mg/kg significantly reduced serum MDA levels in rats.⁷⁵ The *C. japonica* bee pollen ethyl acetate extract (including gallic acid, p-hydroxybenzoic acid, ferulic acid, ellagic acid, quercetin and kaempferol) at doses of 2 and 4 g/kg showed a significant decrease in liver MDA contents in mice liver.⁷⁶ Biflavonoids and kaempferol from shells of *C. oleifera* at a dose of 200 mg/kg reduced levels in blood serum of mice in each group by nearly 50%.⁷⁷ Thus, the strong lipid peroxidation activity of the two *Camellia* species in our study may have been due to the flavonoid or/and phenolic compounds.

Conclusion

Research on polyphenols, lipid peroxidation and cytotoxic activities of two species *C. vuquangensis* and *C. hatinhensis* showed no presence of compounds EGCG, GCG, ECG, and CG, but biological activities of these two species were stronger when compared with previously reported *Camellia* species. The total polyphenol content of the shoots, leaves, and flowers were comparable to that of green tea. Further research is necessary to have evaluate the seasonal variation and the effect of different extraction methods on total polyphenol content to consider them as new sources of tea with economic value. Further studies on structural determination of polyphenol compounds and their biological activities are also needed in the future.

Summary

The total polyphenol contents of leaves, flowers, and young shoots of two yellow *Camellia* species, *C. vuquangensis* and *C. hatinhensis* were comparable to that of green tea.

Extracts of leaves, flowers of *C. vuquangensis* and *C. hatinhensis* showed strong lipid peroxidation inhibitory activities (IC₅₀: 7.92–17.45 µg/mL).

Extracts of leaves and flowers of *C. vuquangensis* and *C. hatinhensis* showed moderate cytotoxic activities against cell lines HepG2, A549, MCF7, SK-Mel-2, HT-29, and AGS (IC₅₀: 34.73–80.58 µg/mL).

Abbreviations

EtOH: Ethanol; **MeOH:** Methanol; **TLC:** Thin-layer chromatography; **HPLC:** High-Performance liquid chromatography; **OD:** Optical density; **SRB:** Sulforhodamine B; **MDA:** Malondialdehyde; **GAE:** Gallic acid equivalent; **HepG2:** Human hepatocellular carcinoma; **A549:** Human lung carcinoma; **MCF7:** Michigan Cancer Foundation-7; **SK-Mel-2:** Human melanoma; **HT-29:** Human colorectal adenocarcinoma; **AGS:** Human gastric adenocarcinoma; **ATP:** Adenosine triphosphate; **ROS:** Reactive oxygen species; **PUFAs:** Polyunsaturated fatty acids; **EGCG:** Epigallocatechin gallate; **GCG:** Galocatechin gallate; **ECG:** Epicatechin gallate; **CG:** Catechin gallate; **NVQ:** Fresh shoots (leaf bud including two youngest leaves of *Camellia vuquangensis*); **LVQ:** Young leaves (third to fifth leaves of *Camellia vuquangensis*); **HVQ:** Flowers of *Camellia vuquangensis*; **NHT:** Fresh shoots (leaf bud including two youngest leaves of *Camellia hatinhensis*); **LHT:** Young leaves (third to fifth leaves of *Camellia hatinhensis*); **HHT:** Flowers of *Camellia hatinhensis*; **TBA:** thiobarbituric acid; **UV:** Ultraviolet; **FBS:** Fetal bovine serum; **EDTA:** Ethylenediaminetetraacetic acid; **DMSO:** Dimethyl sulfoxide; **TCA:** Trichloroacetic acid; **NCI:** National Cancer Institute.

Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Ethical Approval

All experimental procedures that involved animals (mice) were conducted in accordance with the “Guideline for the Care and Use of Laboratory Animals” which was approved by the Medical-Biological Research Ethics Committee of Institute of Biotechnology Scientific Council (No.140/QĐ-CNSH dated April 13, 2011), Vietnam.

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Informed Consent

There are no human subjects in this article and informed consent is not applicable.

References

1. Gaschler MM and Stockwell BR. Lipid peroxidation in cell death. *Biochem Biophys Res Commun* 2017; 482(3): 419–25. DOI: 10.1016/j.bbrc.2016.10.086, PMID 28212725.
2. Desai SN, Farris FF, and Ray SD. Lipid peroxidation. In: Wexler P (ed) *Encyclopedia of Toxicology*. Elsevier, 2014, pp. 89–93.
3. Cai Z. Lipid peroxidation. In: Wexler P (ed) *Encyclopedia of Toxicology*. Elsevier, 2005, pp. 730–34.
4. WHO. *Latest Global Cancer Data: Cancer Burden Rises to 18.1 Million New Cases and 9.6 Million Cancer Deaths in 2018*. International Agency for Research on Cancer. Geneva: World Health Organization, 2018.

5. Comşa Ş, Cîmpean AM, and Raica M. The story of MCF-7 breast cancer cell line: 40 years of experience in research. *Anticancer Res* 2015; 35(6): 3147–54. PMID 26026074.
6. WHO.int [Internet]. *World Health Organization*, updated March 26, 2021. (cited Jun 4, 2021). <https://www.who.int/news-room/fact-sheets/detail/breast-cancer>
7. Ji Y, Cao Y, and Song Y. Green synthesis of gold nanoparticles using a *Cordyceps militaris* extract and their anti-proliferative effect in liver cancer cells (HepG2). *Artif Cells Nanomed Biotechnol* 2019; 47(1): 2737–45. DOI: 10.1080/21691401.2019.1629952, PMID 31304798.
8. WHO [Internet]. *World Health Organization*, April 3, 2003. (cited Jun 4, 2021). <https://www.who.int/news/item/03-04-2003-global-cancer-rates-could-increase-by-50-to-15-million-by-2020>
9. Yin L, Jiang LP, Shen QS, et al. NCAH plays important roles in human colon cancer. *Cell Death Dis* 2017; 8(3): e2680. DOI: 10.1038/cddis.2017.88, PMID 28300828.
10. Smyth EC, Nilsson M, Grabsch HI, et al. Gastric cancer. *Lancet* 2020; 396(10251): 635–48. DOI: 10.1016/S0140-6736(20)31288-5, PMID 32861308.
11. WHO [Internet]. *World Health Organization*, updated July 22, 2002. (cited Jun 4, 2021). <https://www.who.int/news/item/22-07-2002-helping-people-reduce-their-risks-of-skin-cancer-and-cataract>.
12. Yang C, Liu X, Chen Z, et al. Comparison of oil content and fatty acid profile of ten new *Camellia oleifera* Cultivars. *J Lipids* 2016; 2016: 3982486. DOI: 10.1155/2016/3982486, PMID 26942012.
13. Teixeira AM, and Sousa C. A review on the biological activity of *Camellia* species. *Molecules* 2021; 26(8): 2178. DOI: 10.3390/molecules26082178, PMID 33918918.
14. Usman H, Abdulrahman FI, and Usman A. Qualitative phytochemical screening and *in vitro* antimicrobial effects of methanol stem bark extract of *Ficus thonningii* (Moraceae). *Afr J Tradit Complement Altern Med* 2009; 6(3): 289–95. DOI: 10.4314/ajtcam.v6i3.57178, PMID 20448855.
15. Ministry of Health. *Vietnamese Pharmacopoeia V, Part 2*. Hanoi: Medical Publishing House, 2017, pp. 149–51, Appendices.
16. Bijekar S and Gayatri M. Phytochemical profile of *Codiaeum variegatum* (L.) Bl. *Int J Pharm Pharm Sci* 2016; 2(3): 22–31.
17. Alebiosu CO and Yusuf AJ. Phytochemical screening, thin-layer chromatographic studies and UV analysis of extracts of *Citrullus lanatus*. *J Pharm Chem Biol Sci* 2015; 3(2): 214–20.
18. Tang Y, Yu L, Pei Q, et al. Extraction of polysaccharides from *Spirulina* with boiling water. *Agric Sci Technol* 2014; 15(6): 1043–45.
19. Deepa M and Padmaja CK. Preliminary phytochemical analysis and thin layer chromatography of the extracts of *Excoecaria agallocha* L. *Int J Pharm Sci Res* 2014; 5(10): 1000–07.
20. Atcc.org [Internet]. *American Type Culture Collection*. (cited Jun 17, 2022). <https://www.atcc.org/resources/technical-documents/guide-to-subculturing-cell-line-monolayers>
21. Tran LT, Ho DV, Le DV, et al. Three new phloroglucinol derivatives from the aerial parts of *Pogostemon auricularius* and their cytotoxic activity. *Phytochem Lett* 2018; 28: 88–92. DOI: 10.1016/j.phytol.2018.09.016.
22. Stroeve EA and Makarova VG. Determination of lipid peroxidation rate in tissue homogenate laboratory. *Lab Man Biochem* 1989: 243–56.
23. Badmus JA, Adedosu TO, Fatoki JO, et al. Lipid peroxidation inhibition and antiradical activities of some leaf fractions of *Mangifera indica*. *Acta Pol Pharm* 2011; 68(1): 23–9. PMID 21485698.
24. Punyasiri PAN, Jeganathan B, Kottawa-Arachchi JD, et al. Genotypic variation in biochemical compounds of the Sri Lankan tea (*Camellia sinensis* L.) accessions and their relationships to quality and biotic stresses. *J Hort Sci Biotechnol* 2017; 92(5): 502–12. DOI: 10.1080/14620316.2017.1289070.
25. Anesini C, Ferraro GE, and Filip R. Total polyphenol content and antioxidant capacity of commercially available tea (*Camellia sinensis*) in Argentina. *J Agric Food Chem* 2008; 56(19): 9225–29. DOI: 10.1021/jf8022782, PMID 18778031.
26. Zhao C, Li C, Liu S, et al. The galloyl catechins contributing to main antioxidant capacity of tea made from *Camellia sinensis* in China. *Sci World J* 2014; 2014: 863984. DOI: 10.1155/2014/863984, PMID 25243234.
27. Gadkari PV, Kadimi US, and Balaraman M. Catechin concentrates of garden tea leaves (*Camellia sinensis* L.): Extraction/isolation and evaluation of chemical composition. *J Sci Food Agric* 2014; 94(14): 2921–28. DOI: 10.1002/jsfa.6633, PMID 24585505.
28. Ku KM, Choi JN, Kim J, et al. Metabolomics analysis reveals the compositional differences of shade grown tea (*Camellia sinensis* L.). *J Agric Food Chem* 2010; 58(1): 418–26. DOI: 10.1021/jf902929h, PMID 19994861.
29. Ruan J, Haerdter R, and Gerendás J. Impact of nitrogen supply on carbon/nitrogen allocation: A case study on amino acids and catechins in green tea [*Camellia sinensis* (L.) O. Kuntze] plants. *Plant Biol (Stuttg)* 2010; 12(5): 724–34. DOI: 10.1111/j.1438-8677.2009.00288.x, PMID 20701695.
30. Punyasiri PAN, Jeganathan B, Kottawa-Arachchi JD, et al. New sample preparation method for quantification of phenolic compounds of tea (*Camellia sinensis* L. Kuntze): A polyphenol rich plant. *J Anal Methods Chem* 2015; 2015: 964341. DOI: 10.1155/2015/964341, PMID 26543665.
31. Pal S, Ghosh D, Saha C, et al. Total polyphenol content, antioxidant activity and lipid peroxidation inhibition efficacy of branded tea (*Camellia sinensis*) available in India. *Int J Tea Sci* 2012; 8(3): 13–20.
32. Gao X, Lin X, Li X, et al. Cellular antioxidant, methylglyoxal trapping, and anti-inflammatory activities of cocoa tea (*Camellia pitilophylla* Chang). *Food Funct* 2017; 8(8): 2836–46. DOI: 10.1039/c7fo00368d, PMID 28725904.
33. Roy MK, Koide M, Rao TP, et al. ORAC and DPPH assay comparison to assess antioxidant capacity of tea infusions: Relationship between total polyphenol and individual catechin content. *Int J Food Sci Nutr* 2010; 61(2): 109–24. DOI: 10.3109/09637480903292601, PMID 20109129.
34. Zhu C, Zhang S, Fu H, et al. Transcriptome and phytochemical analyses provide new insights into long non-coding RNAs modulating characteristic secondary metabolites of oolong tea (*Camellia sinensis*) in solar-withering. *Front Plant Sci* 2019; 10: 1638. DOI: 10.3389/fpls.2019.01638, PMID 31929782.
35. Bramati L, Aquilano F, and Pietta P. Unfermented rooibos tea: Quantitative characterization of flavonoids by HPLC–UV and

- determination of the total antioxidant activity. *J Agric Food Chem* 2003; 51(25): 7472–74. DOI: 10.1021/jf0347721, PMID 14640601.
36. Li Q, Li J, Liu S, et al. A comparative proteomic analysis of the buds and the young expanding leaves of the tea plant (*Camellia sinensis* L.). *Int J Mol Sci* 2015; 16(12): 14007–38. DOI: 10.3390/ijms160614007.
 37. Tanaka K, Tamaru S, Nishizono S, et al. Hypotriacylglycerolemic and antiobesity properties of a new fermented tea product obtained by tea-rolling processing of third-crop green tea (*Camellia sinensis*) leaves and loquat (*Eriobotrya japonica*) leaves. *Biosci Biotechnol Biochem* 2010; 74(8): 1606–12. DOI: 10.1271/bbb.100197, PMID 20699580.
 38. Wright LP, Mphangwe NI, Nyirenda HE, et al. Analysis of the theaflavin composition in black tea (*Camellia sinensis*) for predicting the quality of tea produced in Central and Southern Africa. *J Sci Food Agric* 2002; 82(5): 517–25. DOI: 10.1002/jsfa.1074.
 39. Wang Y, Wang YM, Lu YT, et al. Influence of different nitrogen sources on carbon and nitrogen metabolism and gene expression in tea plants (*Camellia sinensis* L.). *Plant Physiol Biochem* 2021; 167: 561–66. DOI: 10.1016/j.plaphy.2021.08.034, PMID 34454315.
 40. Yang X, Yu Z, Zhang B, et al. Effect of fluoride on the biosynthesis of catechins in tea [*Camellia sinensis* (L.) O. Kuntze] leaves. *Sci Hortic* 2015; 184: 78–84. DOI: 10.1016/j.scienta.2014.12.031.
 41. Liang-Liang Z, Yong-Mei W, Dong-Mei W, et al. Comparisons of antioxidant activity and total phenolics of *Camellia oleifera* Abel fruit hull from different regions of China. *J Med Plants Res* 2010; 4(14): 1407–13.
 42. Yang R, Guan Y, Wang W, et al. Antioxidant capacity of phenolics in *Camellia nitidissima* Chi flowers and their identification by HPLC Triple TOF MS/MS. *PLoS One* 2018; 13(4): e0195508. DOI: 10.1371/journal.pone.0195508, PMID 29634769.
 43. Zhang X, Feng J, Su S, et al. Hepatoprotective effects of *Camellia nitidissima* aqueous ethanol extract against CCl₄-induced acute liver injury in SD rats related to Nrf2 and NF-κB signalling. *Pharm Biol* 2020; 58(1): 239–46. DOI: 10.1080/13880209.2020.1739719, PMID 32202453.
 44. Zhang HL, Wu QX, Wei X, et al. Pancreatic lipase and cholesterol esterase inhibitory effect of *Camellia nitidissima* Chi flower extracts *in vitro* and *in vivo*. *Food Biosci* 2020; 37: 100682. DOI: 10.1016/j.fbio.2020.100682.
 45. Lin JN, Lin HY, Yang NS, et al. Chemical constituents and anticancer activity of yellow camellias against MDA-MB-231 human breast cancer cells. *J Agric Food Chem* 2013; 61(40): 9638–44. DOI: 10.1021/jf4029877, PMID 24001127.
 46. Song L, Wang X, Zheng X, et al. Polyphenolic antioxidant profiles of yellow camellia. *Food Chem* 2011; 129(2): 351–57. DOI: 10.1016/j.foodchem.2011.04.083, PMID 30634237.
 47. Li K, Shi X, Yang X, et al. Antioxidative activities and the chemical constituents of two Chinese teas, *Camellia kucha* and *C. ptilophylla*. *Int J Food Sci Technol* 2012; 47(5): 1063–71. DOI: 10.1111/j.1365-2621.2012.02942.x.
 48. Samanta S. Potential bioactive components and health promotional benefits of tea (*Camellia sinensis*). *J Am Nutr Assoc* 2022; 41(1): 65–93. DOI: 10.1080/07315724.2020.1827082, PMID 33216711.
 49. Bedrood Z, Rameshrad M, and Hosseinzadeh H. Toxicological effects of *Camellia sinensis* (green tea): A review. *Phytother Res* 2018; 32(7): 1163–80. DOI: 10.1002/ptr.6063, PMID 29575316.
 50. Saeed M, Naveed M, Arif M, et al. Green tea (*Camellia sinensis*) and l-theanine: Medicinal values and beneficial applications in humans—A comprehensive review. *Biomed Pharmacother* 2017; 95: 1260–75. DOI: 10.1016/j.biopha.2017.09.024, PMID: 28938517.
 51. Sinija VR and Mishra HN. Green tea: Health benefits. *J Nutr Environ Med* 2008; 17(4): 232–42. DOI: 10.1080/13590840802518785.
 52. Hughes JP, Rees S, Kalindjian SB, et al. Principles of early drug discovery. *Br J Pharmacol* 2011; 162(6): 1239–49. DOI: 10.1111/j.1476-5381.2010.01127.x, PMID 21091654.
 53. Ramirez-Mares MV, Chandra S, De Mejia EG. *In vitro* chemopreventive activity of *Camellia sinensis*, *Ilex paraguariensis* and *Ardisia compressa* tea extracts and selected polyphenols. *Mutat Res* 2004; 554(1–2): 53–65. DOI: 10.1016/j.mrfmmm.2004.03.002, PMID 15450404.
 54. Lu QY, Yang Y, Jin YS, et al. Effects of green tea extract on lung cancer A549 cells: Proteomic identification of proteins associated with cell migration. *Proteomics* 2009; 9(3): 757–67. DOI: 10.1002/pmic.200800019, PMID 19137550.
 55. Costa D, Santos R, Andrade EDS, et al. Green tea extract (*Camellia sinensis*) as a potential antitumoral agent on breast cancer cells (FS13-04-19). *Curr Dev Nutr* 2019; 3(Suppl_1): 409. DOI: 10.1093/cdn/nzz030.FS13-04-19.
 56. Hajiaghaalipour F, Kanthimathi MS, Sanusi J, et al. White tea (*Camellia sinensis*) inhibits proliferation of the colon cancer cell line, HT-29, activates caspases and protects DNA of normal cells against oxidative damage. *Food Chem* 2015; 169: 401–10. DOI: 10.1016/j.foodchem.2014.07.005, PMID 25236244.
 57. Hashemi L, Asadi-Samani M, Moradi MT, et al. *In vitro* anti proliferative activity, antioxidant potential and total phenolic compounds of black tea extract. *Int J Pharm* 2017; 7(2): 19–25.
 58. Zhang L, Santos JS, Cruz TM, et al. Multivariate effects of Chinese keemun black tea grades (*Camellia sinensis* var. *sinensis*) on the phenolic composition, antioxidant, antihemolytic and cytotoxic/cytoprotection activities. *Food Res Int* 2019; 125: 108516. DOI: 10.1016/j.foodres.2019.108516, PMID 31554085.
 59. He D, Zhang P, Sai X, et al. *Camellia euphlebica* flower extract inhibits oleic acid-induced lipid accumulation via reduction of lipogenesis in HepG2 cells. *Eur J Integr Med* 2018; 17: 1–8. DOI: 10.1016/j.eujim.2017.11.002.
 60. Yang XR, Wang YY, La KK, et al. Inhibitory effects of cocoa tea (*Camellia ptilophylla*) in human hepatocellular carcinoma HepG2 *in vitro* and *in vivo* through apoptosis. *J Nutr Biochem* 2012; 23(9): 1051–57. DOI: 10.1016/j.jnutbio.2011.05.010, PMID 22018604.
 61. Santos JS, Deolindo CTP, Hoffmann JF, et al. Optimized *Camellia sinensis* var. *sinensis*, *Ilex paraguariensis*, and *Aspalathus linearis* blend presents high antioxidant and anti-proliferative activities in a beverage model. *Food Chem* 2018;

- 254: 348–58. DOI: 10.1016/j.foodchem.2018.02.021, PMID 29548463.
62. Chachiyo S, Kulprachakarn K, Saenjum C, et al. Toxicity evaluation of *Camellia sinensis* var. *assamica* and its fermented miang product. *Pharmacog Res* 2020; 12(4): 430. DOI: 10.4103/pr.pr_22_20.
63. Way TD, Lin HY, Hua KT, et al. Beneficial effects of different tea flowers against human breast cancer MCF-7 cells. *Food Chem* 2009; 114(4): 1231–36. DOI: 10.1016/j.foodchem.2008.10.084.
64. Ngo DQ, Nguyen TC, Nguyen TD, et al. A novel biopolymer nano-complex based on fish scale collagen, konjac glucomannan, *Camellia chrysantha* polyphenols and ginsenoside Rb1: Preparation, characterization and its bioactivity. *J Polym Environ* 2021; 29(7): 2150–63. DOI: 10.1007/s10924-020-02022-0.
65. Li-na J, Ji-yuan L, Zheng-qi F, et al. Content analysis of polyphenols in flowers of yellow *Camellia*. *For Res* 2020; 33(4): 117–26.
66. Peng L, Wang X, Shi X, et al. Characterization of the constituents and antioxidative activity of cocoa tea (*Camellia ptillophylla*). *Food Chem* 2011; 129(4): 1475–82. DOI: 10.1016/j.foodchem.2011.05.120.
67. Wang CC, Ho CT, Lee SC, et al. Isolation of eugenyl β -primeveroside from *Camellia sasanqua* and its anticancer activity in PC3 prostate cancer cells. *J Food Drug Anal* 2016; 24(1): 105–11. DOI: 10.1016/j.jfda.2015.06.005, PMID 28911392.
68. Zhu LF, Xu M, Zhu HT, et al. New flavan-3-ol dimer from green tea produced from *Camellia taliensis* in the Ai-Lao mountains of Southwest China. *J Agric Food Chem* 2012; 60(49): 12170–76. DOI: 10.1021/jf302726t, PMID 23167720.
69. Tsujita T, Matsuo Y, Saito Y, et al. Enzymatic oxidation of ellagitannin and a new ellagitannin metabolite from *Camellia japonica* leaves. *Tetrahedron* 2017; 73(5): 500–07. DOI: 10.1016/j.tet.2016.12.027.
70. Ostrowska J, Łuczaj W, Kasacka I, et al. Green tea protects against ethanol-induced lipid peroxidation in rat organs. *Alcohol* 2004; 32(1): 25–32. DOI: 10.1016/j.alcohol.2003.11.001, PMID 15066700.
71. Burckhardt IC, Gozal D, Dayyat E, et al. Green tea catechin polyphenols attenuate behavioral and oxidative responses to intermittent hypoxia. *Am J Respir Crit Care Med* 2008; 177(10): 1135–41. DOI: 10.1164/rccm.200701-110OC, PMID 18276944.
72. Lu W and Wen J. Neuroprotective roles of total flavones of *Camellia* on early brain injury and cognitive dysfunction following subarachnoid hemorrhage in rats. *Metab Brain Dis* 2020; 35(5): 775–83. DOI: 10.1007/s11011-020-00567-6, PMID 32219683.
73. Huang HL, Lin CC, Jeng KC, et al. Fresh green tea and gallic acid ameliorate oxidative stress in kainic acid-induced status epilepticus. *J Agric Food Chem* 2012; 60(9): 2328–36. DOI: 10.1021/jf203709q, PMID 22324774.
74. Haidari F, Omidian K, Raffei H, et al. Green tea (*Camellia sinensis*) supplementation to diabetic rats improves serum and hepatic oxidative stress markers. *Iran J Pharm Res* 2013; 12(1): 109–14. PMID 24250578.
75. Lu W, Xv L, and Wen J. Protective effect of extract of the *Camellia japonica* L. on cerebral ischemia-reperfusion injury in rats. *Arq Neuropsiquiatr* 2019; 77(1): 39–46. DOI: 10.1590/0004-282x20180146.
76. Xu Y, Cao X, Zhao H, et al. Impact of *Camellia japonica* bee pollen polyphenols on hyperuricemia and gut microbiota in potassium oxonate-induced mice. *Nutrients* 2021; 13(8): 2665. DOI: 10.3390/nu13082665, PMID 34444825.
77. Ye Y, Guo Y, Luo YT, et al. Isolation and free radical scavenging activities of a novel biflavonoid from the shells of *Camellia oleifera* Abel. *Fitoterapia* 2012; 83(8): 1585–89. DOI: 10.1016/j.fitote.2012.09.006, PMID 22982330.